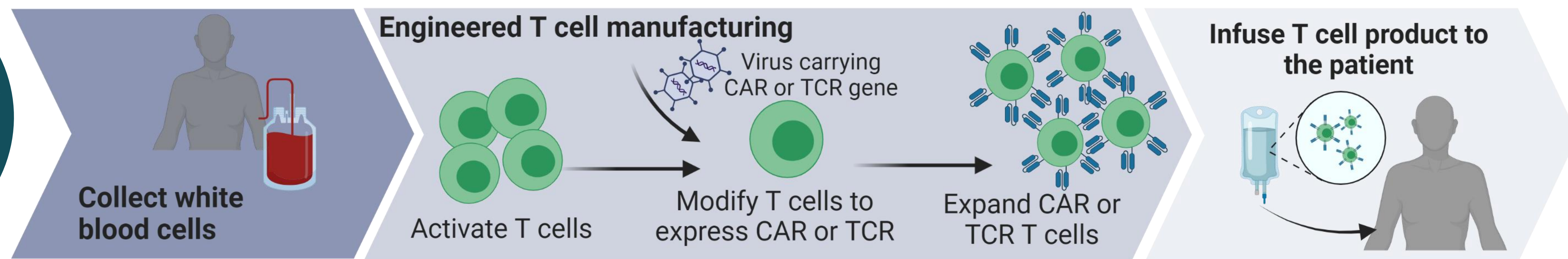


CellFit: T-cells fit to fight cancer

Léa Rosselle¹, Fatemeh Kaveh¹, Hanne Haslene-Hox², Hanne Hein Trøen², Maxi-Lu Böschén³, Tuva Holt Hereng³, Evan Zynda³, Sébastien Wälchli¹, Else Marit Inderberg¹.

The use of T lymphocytes in adoptive cell therapy (ACT) shows great promise for treatment of cancers otherwise incurable. However, one of the largest challenges faced in cell-based cancer therapy is to provide an efficient and scalable production. The use of “living drugs” requires precise logistics at all stages of cell life: development, manufacturing, transport and finally the infusion to the patient. With the CellFit project, we aim to define optimal growth conditions for improved manufacturing of adapted therapeutic T cells required for solid tumor treatment.

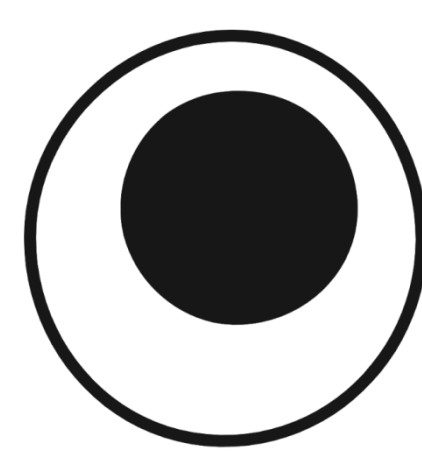
ADOPTIVE CELL THERAPY AGAINST CANCER



CURRENTLY

- ✗ Heterogeneous and exhausted population of T cells
- ✗ Lack of long-term persistence
- ✗ Loss of efficiency after injection

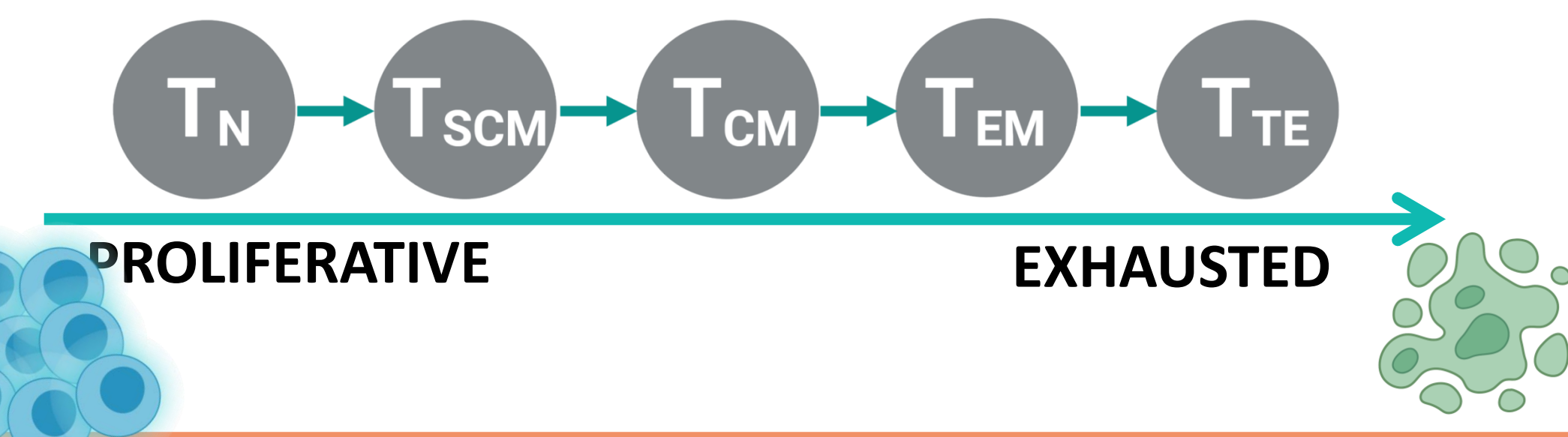
HOW DO WE SOLVE THESE ISSUES ?



STEM-CELL LIKE T CELL FOR A POWERFUL CULTURE

The stem cell state has been studied for many years for their higher potential to self renew in many tissue. In the case of T cell, Stem cell memory T cells (T_{scm}) are able to differentiate into T-cell central memory (T_{cm}) and T-cell effector memory (T_{em}) and show low signs of exhaustion. This cell subset is a good candidate for improving T-cell therapy but is present in very low numbers in T-cell culture. To be able to produce T cells of young/memory phenotype, a controlled T-cell activation process is needed. Many different serum sources, cytokines and growth factors or chemicals as well as stimulation can be explored to get an optimal T-cell culture.

T-CELL HETEROGENEITY

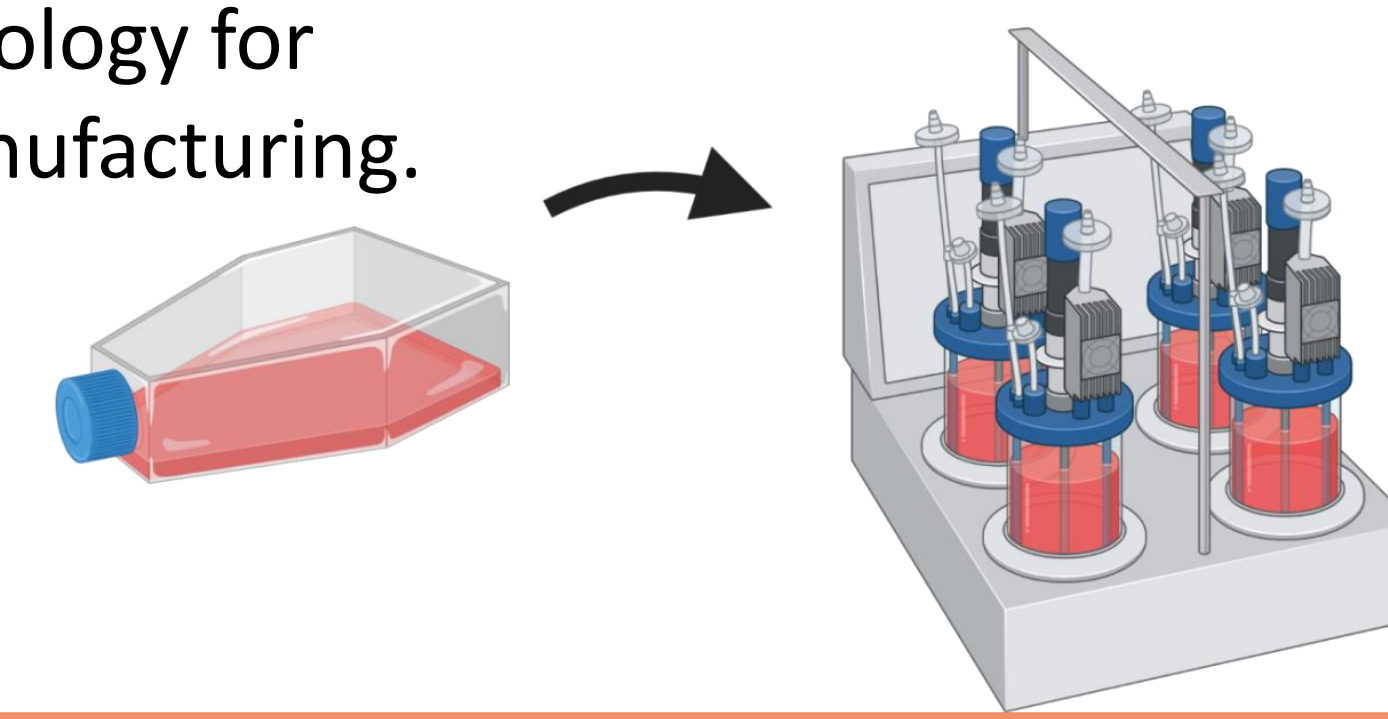


INNOVATIVE REAGENT DEVELOPMENT

The design of new media and Dynabeads for T-cell expansion according to clinical grade cell culture exigencies is a key part of the optimisation of T-cell manufacturing.

LARGE SCALE AND AUTOMATED CULTURE

After optimal culture selection, we will be able to establish a novel manufacturing method adapted to large-scale cGMP production and thus provide novel production methodology and technology for T-cell manufacturing.



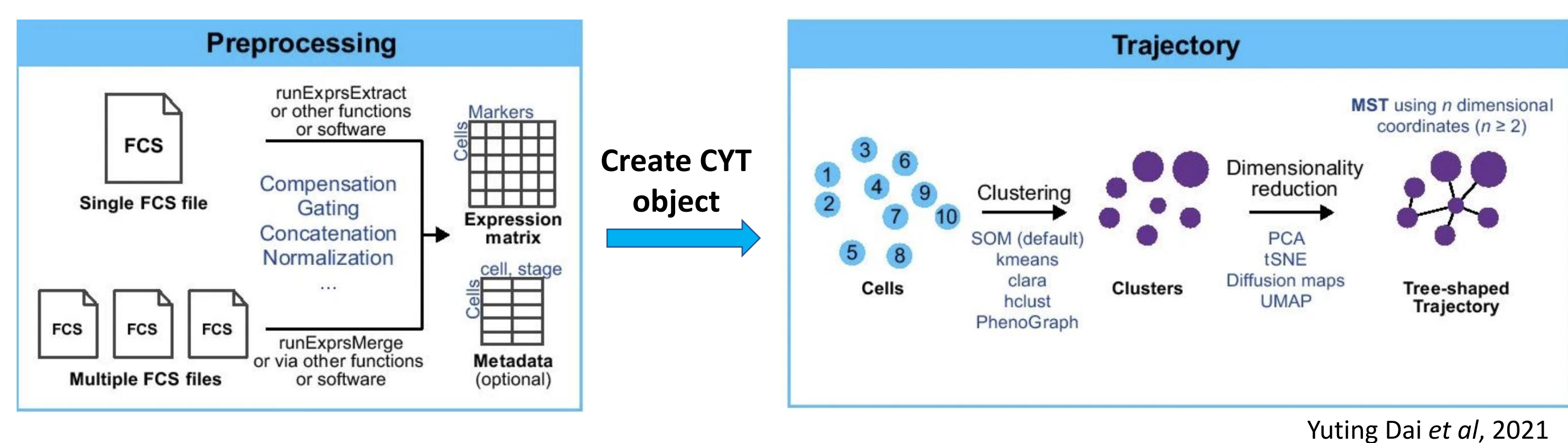
CULTURE OPTIMISATION

CELLFIT PROJECT

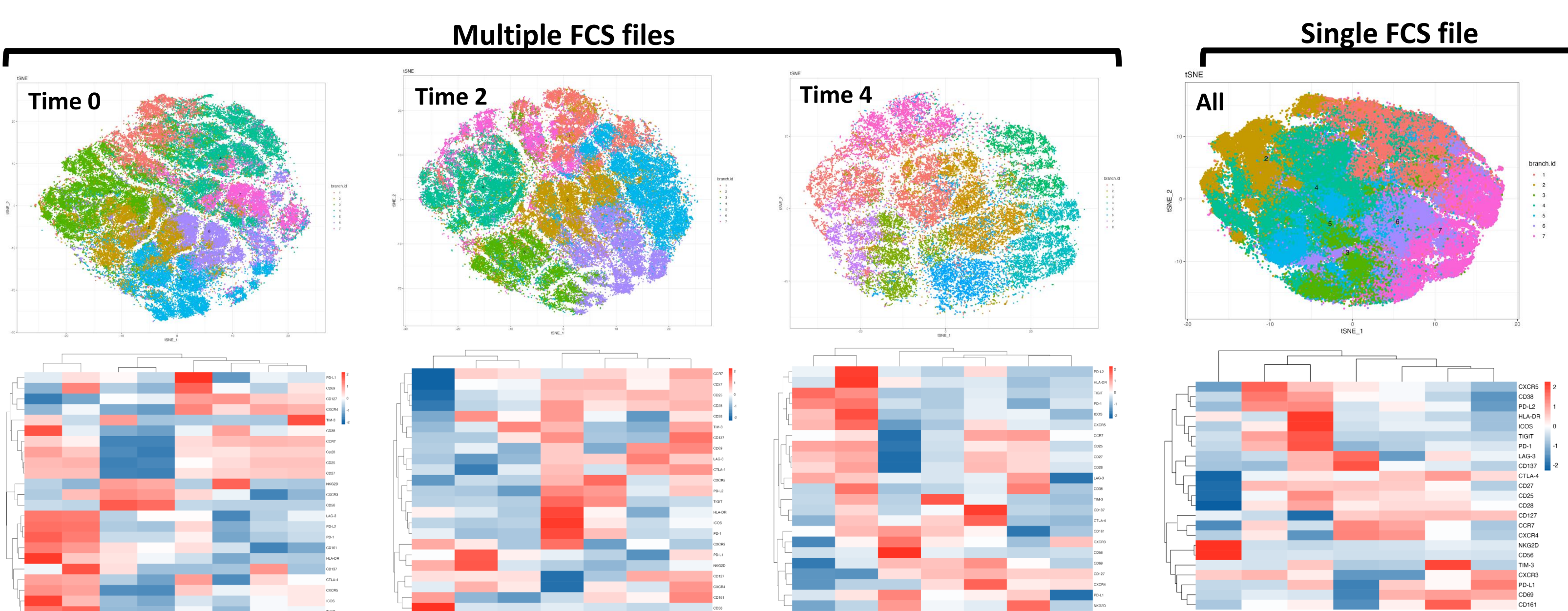
BIOINFORMATICS

PIPELINE

The bioinformatics pipeline for mass cytometry data consists of below R-packages: **Pre-processing:** [CATALYST](#) which will be used only for concatenations of truncated fcs.files and quality control. **Analysis:** will be done through CytoTree & FlowSOM packages. We will test 5 types of clustering methods through CytoTree and compare results to the commonly used Cytobank.



Patient comparison: Analysis of blood within 3 time points (0, 2, 4 months & all together) from a breast cancer patient treated with two different drugs.

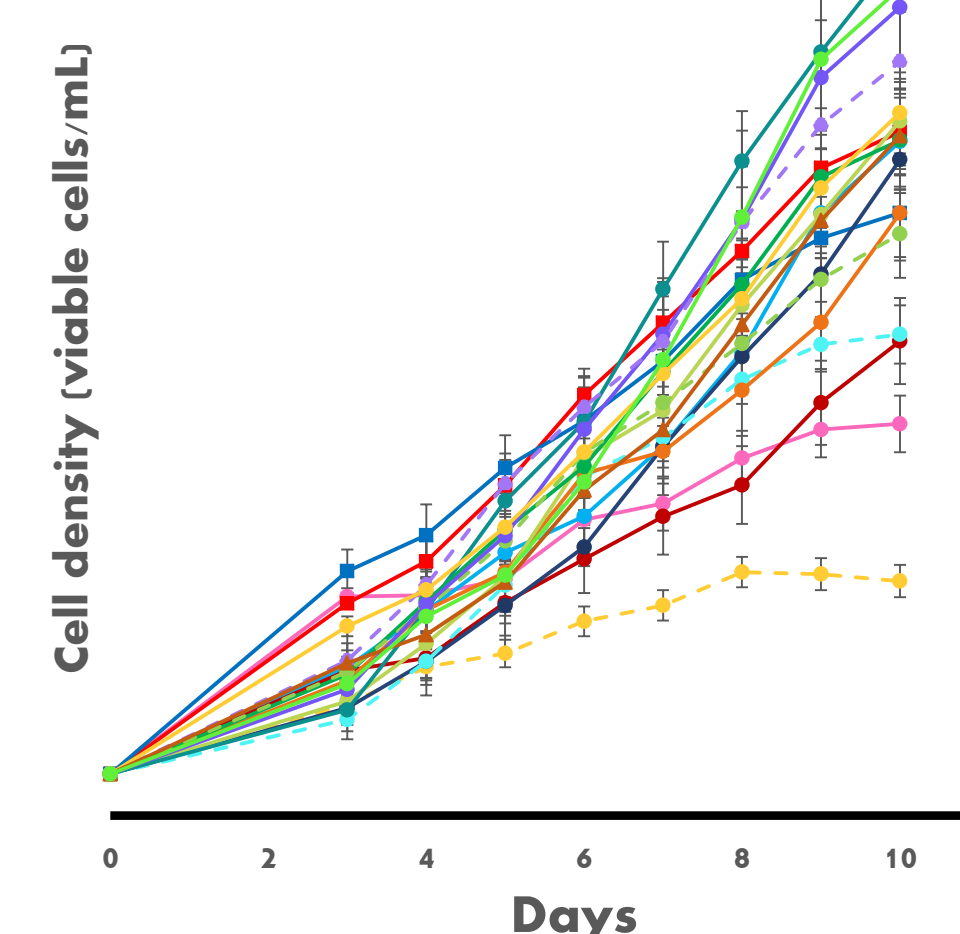


T-CELL POPULATION STUDY

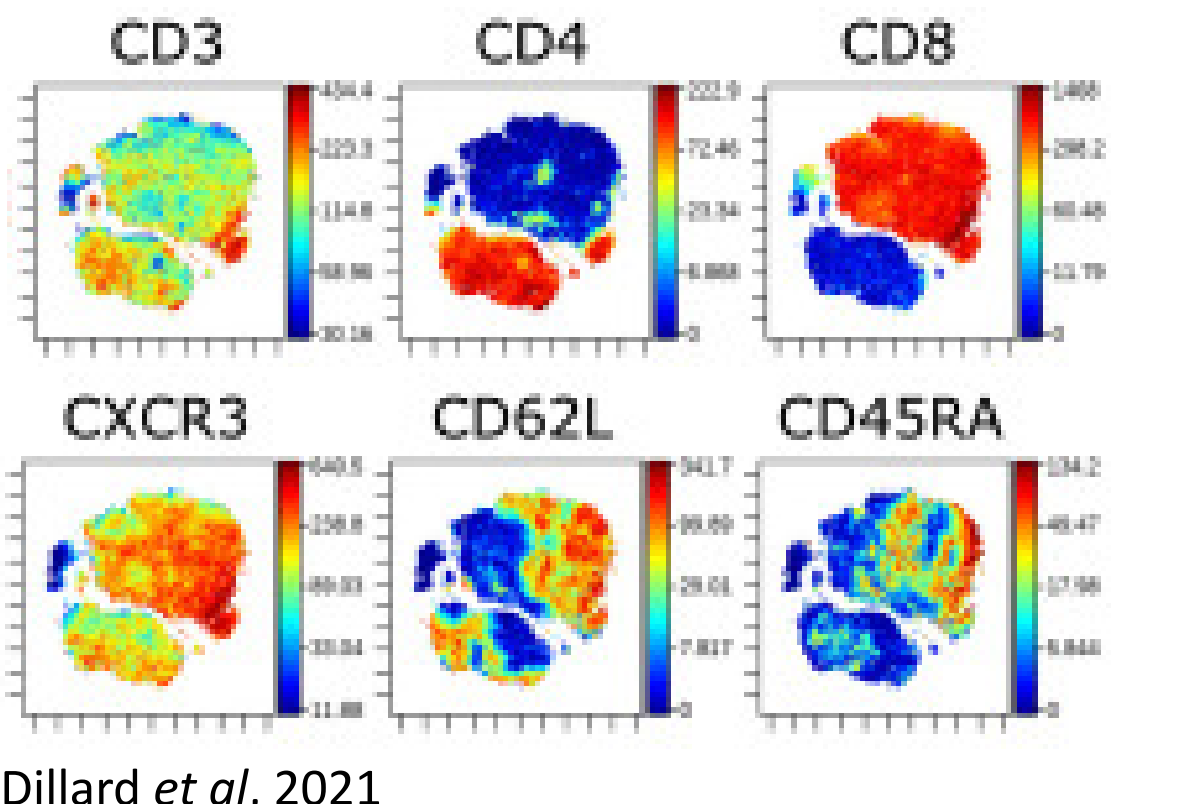
HIGH THROUGHPUT SCREENING

The set-up of T-cell expansion in 96-well plate format allow the extensive screening of culture conditions. The T-cell culture methods will be translated from manual to robotic set-ups where all pipetting, cell culturing and assay read-outs will be performed automatically with robots coupled to incubators, spectrophotometer and high-content confocal microscope.

T-cell growth under different cultivation conditions can be tested with many conditions in parallel

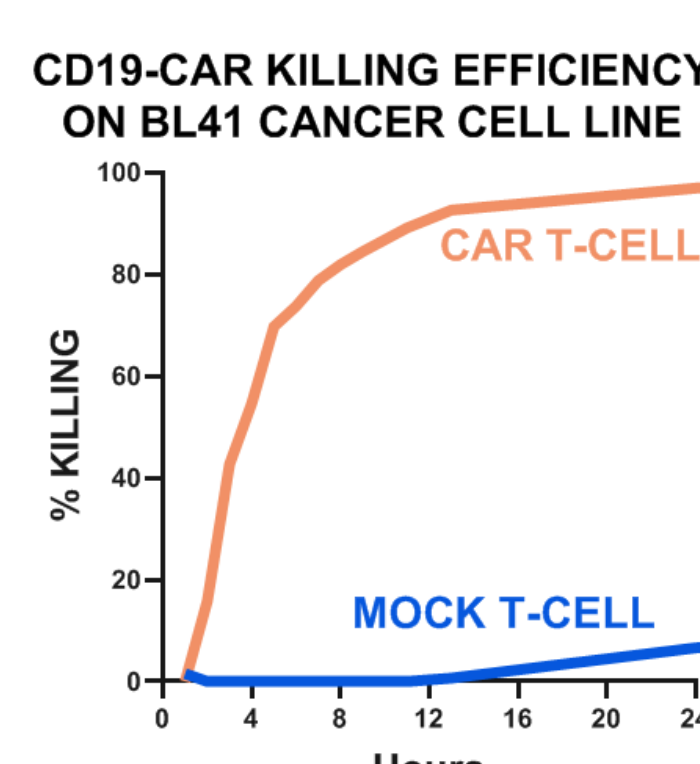


FLOW AND MASS CYTOMETRY

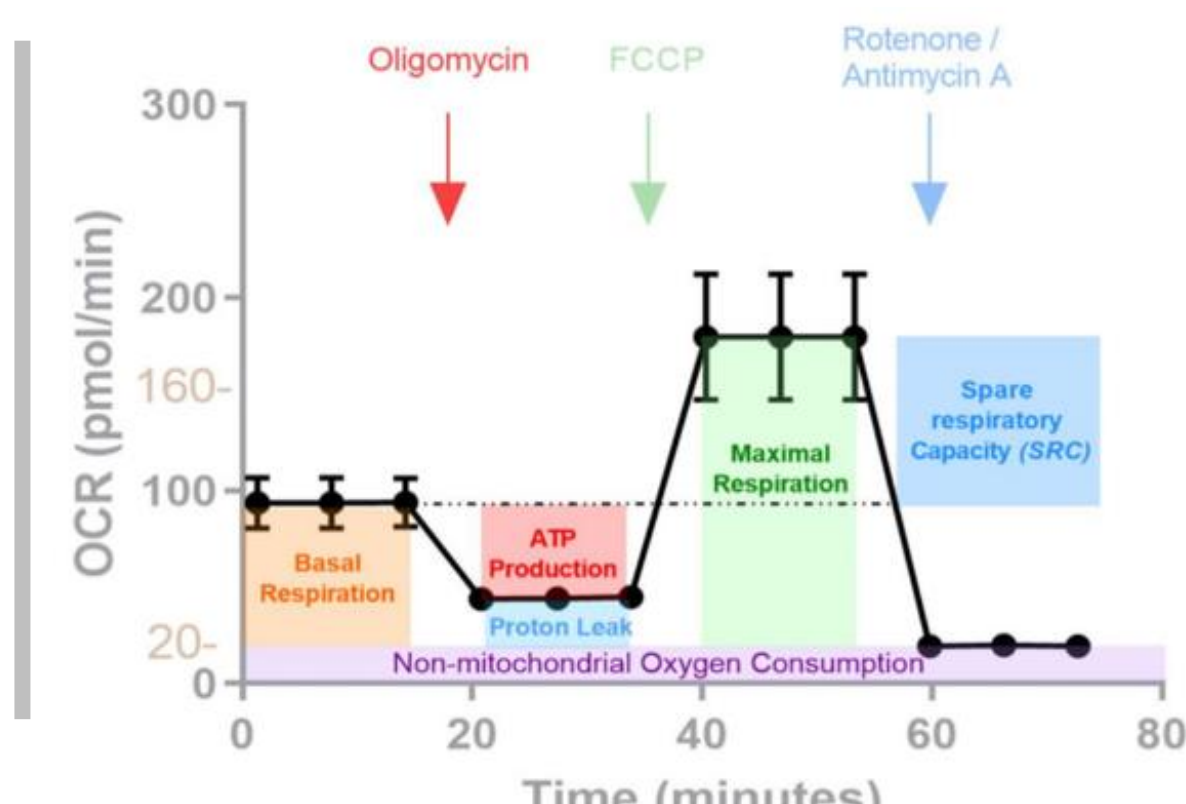


Dillard et al, 2021

FUNCTIONAL ASSAYS



METABOLIC ANALYSIS



Forcados et al, 2022

Evaluate phenotype and metabolism of cellular subsets as well as therapeutic effect of produced cells.